

Evaluation of Antimicrobial Resistance and Bacterial Spectrum of Infections at a Tertiary Care Teaching Hospital: A Retrospective Analysis

Chaman Lal Girdhar^{1*}, Naresh Jyoti²

^{1*}Assistant Professor, Department of Microbiology,

²Associate Professor, Department of Pharmacology,

Chintpurni Medical College & Hospital, Bungal, Pathankot, Punjab, India.

Article History

Received: 16 Oct 2015

Revised: 03 Nov 2015

Accepted: 18 Nov 2015

*Correspondence to:

Dr. C L Girdhar
Assistant Professor,
Department of
Microbiology,
Chintpurni Medical
College & Hospital,
Bungal, Pathankot,
Punjab, India.

ABSTRACT

Introduction: It is essential to study the antimicrobial resistance and spectra of activity of bacteria time to time to provide data for the rational use of antibiotics, henceforth the present retrospective study was taken into consideration.

Materials and Methods: The present study was conducted over total 500 samples of bacterial culture that comprised of blood samples, swabs from ear, nose, throat, skin and wound infections and urine samples. Bacterial isolates were identified using standard laboratory methods. Identification of isolates was confirmed by PCR. Data so obtained was analyzed using the SPSS Version 17 software and was arranged according to characteristics and represented as a number and percentage.

Results: Staphylococcus aureus was found in 36% cases, Enterococcus faecalis in 26%, Escherichia coli 18%, Streptococcus pneumonia in 11%, Salmonella enteric in 5% cases, Klebsiella pneumonia in 9%, Pseudomonas aeruginosa in 6%, Salmonella enteric in 5%, Enterococcus faecium in 5%, and Proteus mirabilis in 4% cases. 38% were Methicillin-resistant S aureus.

Conclusion: The bacterial spectrum of infection and antimicrobial resistance of various strains should be evaluated time to time to monitor antibiotic resistance as well as prevalence of multidrug resistant strain as it is important to plan treatment strategy.

KEYWORDS: Antibiotic resistance, Methicillin-resistant Staphylococcus aureus, Microbiology.

INTRODUCTION

Antimicrobial resistance is not a novel concern, however the number of resistant organisms, the geographic locations affected by drug resistance, and the breadth of resistance in single organisms are unparalleled and mounting.¹ The knowledge of the infection epidemiology and the antibiotic resistance patterns is important to guide optimal empiric treatment in critically ill patients.² Diseases and disease agents that were once thought to be controlled by antibiotics are returning in new leagues resistant to these therapies.¹ In view of this, it is essential to study the antimicrobial resistance and spectra of activity of bacteria time to time to provide data for the rational use of antibiotics, henceforth the present retrospective study was taken into consideration.

MATERIALS AND METHODS

The present retrospective study was conducted in Department of Microbiology, Chintpurni Medical College & Hospital, Bungal, Pathankot, Punjab (India) over total 500 samples of bacterial culture that comprised of blood samples, swabs from ear, nose, throat, skin and wound infections and urine samples. Samples were randomly selected from the data of patients with age from 18 to 50 years from microbiological laboratory over a period of one year. Ethical clearance was taken for the commencement of the study.

Commercial blood cultures were used to study blood infections. Sterile cotton swabs were used for the collection of samples from ear, nose, throat, skin and

wound infections. Sterile single-use pots were used for collection of urine samples for microbiological culture. Bacterial isolates were identified using standard laboratory methods. Identification of isolates was confirmed by PCR.

Disk-diffusion method was used for antimicrobial susceptibility testing as described according to guidelines for laboratory detection of Clinical and

Laboratory Standards Institute (CLSI). PBP2a-agglutination test was used for assessing methicillin-resistance for all ceftoxitin-resistant *Staphylococcus aureus*.

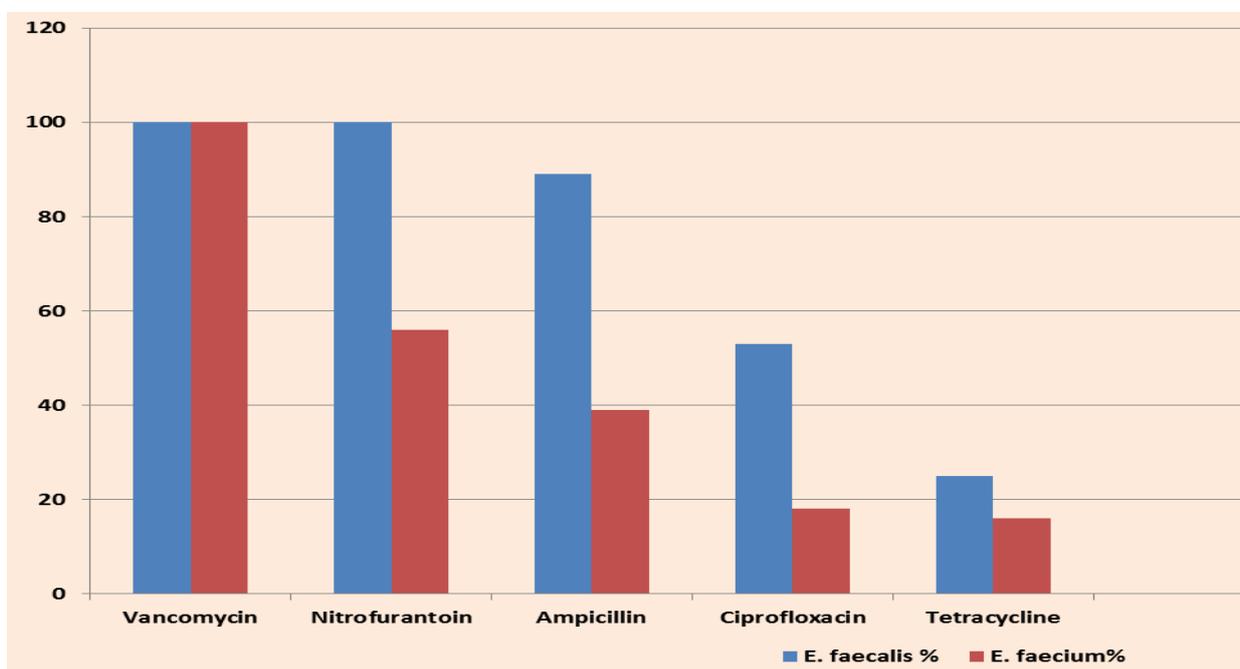
Data so obtained was analyzed using the SPSS Version 17 software and was arranged according to characteristics and represented as a number and percentage.

Table 1: Bacterial spectrum of infection in present study.

Pathogen	Patients n (%)
<i>Staphylococcus aureus</i>	36%
<i>Salmonella enteric</i>	5%
<i>Streptococcus pneumonia</i>	11%
<i>Enterococcus faecalis</i>	26%
<i>Enterococcus faecium</i>	5%
<i>Escherichia coli</i>	18%
<i>Klebsiella pneumoniae</i>	9%
<i>Proteus mirabilis</i>	4%
<i>Pseudomonas aeruginosa</i>	6%

Table 2: Resistance rate of Methicillin resistant *Staphylococcus aureus* to various antibiotics

<i>Staphylococcus aureus</i>	Antibiotics	Resistance rate
Methicillin resistant <i>Staphylococcus aureus</i>	Methicillin	38%
	Erythromycin	100%
	Fluoroquinolones	62%
	Clindamycin	71%



Graph 1: Antibiotic sensitivity of *Enterococcus faecalis* and *Enterococcus faecium*

RESULTS

Staphylococcus aureus was found in 36% cases, *Enterococcus faecalis* in 26%, *Escherichia coli* 18%, *Streptococcus pneumoniae* in 11%, *Salmonella enteric* in 5% cases, *Klebsiella pneumoniae* in 9%, *Pseudomonas aeruginosa* in 6%, *Enterococcus faecium* in 5%, and *Proteus mirabilis* in 4% cases (table 1).

From the *Staphylococcus aureus* isolates, 38% were Methicillin-resistant *S. aureus* (MRSA) with 100% of them showed resistance to erythromycin, 62 % fluoroquinolones and 71% to clindamycin (table 2).

All isolated strains of *E. faecalis* were sensitive to vancomycin and to nitrofurantoin, 89% strains were sensitive to ampicillin, 53% to ciprofloxacin and 25% to tetracycline. All strains of *E. faecium* were sensitive to vancomycin, 56% to nitrofurantoin, 39% were sensitive to ampicillin, 18% to ciprofloxacin and 16% to tetracycline.

DISCUSSION

The most prevalent pathogen found in present study was *Staphylococcus aureus* followed by *Enterococcus faecalis* and *Escherichia coli*. Similarly, Alabi AS et al² conducted a study retrospective analysis of microbiological findings from a secondary care hospital in South Africa and reported *Staphylococcus aureus* as the most prevalent. Orsini J et al³ conducted a prospective observational study among adult patients with clinical signs of sepsis in hospital in New York and reported that *Staphylococcus aureus* was found in 20.8% patients, *Enterococcus spp* in 16.6% and *Streptococcus pneumoniae* in 11.1%.

Staphylococcus aureus is one of the most considerable human pathogens responsible for nosocomial and community-acquired infections. It can cause a range of infectious diseases from mild conditions, such as soft tissue infections, to severe life-threatening debilitation, such as endocarditis.⁴ Methicillin-resistant *Staphylococcus aureus* is a major pathogen worldwide; MRSA infections are associated with increased morbidity and mortality, in comparison with other *S. aureus* infections. Over the past decade, the changing pattern of resistance in *S. aureus* has underscored the need for new antimicrobial agents.⁵

In the present study, from the *Staphylococcus aureus* isolates, 38% were Methicillin-resistant *S. aureus*. Chapin KC et al⁶ conducted a study to identify Methicillin-Resistant *Staphylococcus aureus* from blood cultures and found that of the 70 blood cultures positive for *S. aureus*, 44 had methicillin-resistant *S. aureus* and 26 had methicillin-susceptible *S. aureus* as determined by the oxacillin screen agar method using isolated colonies. Shitu AO et al⁷ studied antibiotic resistance and molecular epidemiology of *Staphylococcus aureus* in Nigeria and reported that *ermA* gene was identified in all erythromycin-resistant MRSA isolates, while two

erythromycin-resistant MSSA isolates possessed the *msrA* gene. All the gentamicin-resistant isolates carried the *aacA-aphD* gene. Methicillin resistance has become an important problem among hospital isolates as these strains are also often resistant to other antibiotics such as aminoglycosides, tetracyclines and quinolones.⁴ Methicillin resistance in *S. aureus* involves an altered target site due to an acquired penicillin-binding protein (PBP 2a) with decreased affinity to β -lactams. The *mecA* gene encodes this protein and is located on a mobile SCCmec cassette chromosome. This genetic element confers resistance to most currently available β -lactam antibiotics.⁸ Zahid KF et al⁹ evaluated bacterial spectrum and susceptibility patterns of pathogens in adult febrile neutropenic patients and reported that 33% strains of *Staphylococcus aureus* were methicillin resistant and *Escherichia coli* was the pathogen found in 23.1% of patients, followed by *Staphylococcus epidermidis* in 13.9%, *Pseudomonas aeruginosa* in 12.5% and *Staphylococcus aureus* in 7.9%.

The importance of the genus *Enterococcus* clinically is directly related to its antibiotic resistance, which contributes to the risk of colonization and infection. The species of the greatest clinical importance are *Enterococcus faecalis* and *Enterococcus faecium*.¹⁰ Enterococci show natural resistance to many antibiotics as well as also acquire easy resistance to antibiotics. Infections caused by multiresistant strains are difficult to treat, are chronic and recurrent as well as are fatal sometimes.¹¹

Enterococcus faecalis, while normally a gut commensal, is a frequent cause of many serious human infections, including urinary tract infections, endocarditis, bacteremia, and wound infections. Among the diseases that *E. faecalis* causes, urinary tract infections are the most common, responsible for approximately 110,000 cases yearly, many of which are nosocomial. Infections with *E. faecalis* can be especially troublesome to treat because of their frequent resistance to multiple antibiotics, including vancomycin, a drug of last resort for many gram-positive infections.¹² However, in the present study, all isolated strains of *E. faecalis* were sensitive to vancomycin and to nitrofurantoin, 89% strains were sensitive to ampicillin, 53% to ciprofloxacin and 25% to tetracycline. All strains of *E. faecium* were sensitive to vancomycin, 56% to nitrofurantoin, 39% were sensitive to ampicillin, 18% to ciprofloxacin and 16% to tetracycline.

A major cause behind the survival of these organisms in hospital environment is the intrinsic resistance to several commonly used antibiotics and, possibly more importantly, their ability to acquire resistance to all presently available antibiotics, either by receipt of foreign genetic material through the transfer of plasmids and transposons or by mutation. The emergence of vancomycin-resistant enterococci is a matter of concern,

as it is very difficult to control when once established. Moreover, there can be transfer of resistant gene from enterococci to *Staphylococcus aureus* thereby posing a threat to the patient safety and also challenges for the treating physicians.¹³

CONCLUSION

The present study concludes that bacterial spectrum of infection and antimicrobial resistance of various strains should be evaluated time to time to monitor antibiotic resistance as well as prevalence of multidrug resistant strain. It is also of importance as epidemiological profile varies place to place and the knowledge of the infection epidemiology and the antibiotic resistance patterns is important to plan treatment strategy.

REFERENCES

1. Levy SB, Marshall B. Antibacterial resistance worldwide: causes, challenges and responses. *Nature Medicine Supplement* 2004;10(12):122-9.
2. Alabi AS, Frielinghaus L, Kaba H, Kusters K, Huson MAM, Kahl BC, et al. Retrospective analysis of antimicrobial resistance and bacterial spectrum of infection in Gabon, Central Africa. *BMC Infectious Diseases* 2013; 13:455.
3. Orsinia J, Mainardia C, Muzyloa E, Karkia N, Cohenb N, Sakoulas G. Microbiological Profile of Organisms Causing Bloodstream Infection in Critically Ill Patients. *J Clin Med Res* 2012;4(6):371-7.
4. Akcama FZ, Tinazb BG, Kayaa O, Tiglia A, Tureb E, Hosogluc S. Evaluation of methicillin resistance by cefoxitin disk diffusion and PBP2a latex agglutination test in *mecA*-positive *Staphylococcus aureus*, and comparison of *mecA* with *femA*, *femB*, *femX* positivities. *Microbiological Research* 2009; 164; 400-403.
5. Appelbaum PC. Microbiology of Antibiotic Resistance in *Staphylococcus aureus*. *Clinical Infectious Diseases* 2007; 45:165-70.
6. Chapin KC, Musgnug MC. Evaluation of Penicillin Binding Protein 2a Latex Agglutination Assay for Identification of Methicillin-Resistant *Staphylococcus aureus* Directly from Blood Cultures. *Journal of Clinical Microbiology* 2004; 42(3):1283-4.
7. Shittu AO, Okon K, Adesida S, Oyedara O, Witte W, Strommenger B, Layer F, Nubel U. Antibiotic resistance and molecular epidemiology of *Staphylococcus aureus* in Nigeria. *Microbiology* 2011; 11:92.
8. Stapleton PD, Taylor PW. Methicillin resistance in *Staphylococcus aureus*: mechanisms and modulation. *Science progress* 2002; 85(1):57-72.
9. Zahid KF, Hafeez H, Afzal A. Bacterial spectrum and susceptibility patterns of pathogens in adult febrile neutropenic patients: a comparison between two time periods. *J Ayub Med Coll Abbottabad* 2009; 21(4):146-9.
10. Kristich CJ, Rice LB, Arias CA. In: Gilmore MS, Clewell DB, Ike Y, Shankar N. *Enterococci: From Commensals to Leading Causes of Drug Resistant Infection*. Boston: Massachusetts Eye and Ear Infirmary; 2014.
11. Rudy M, Nowakowska M, Wiechula B, Zientara M, Radosz-Komoniewska H. Antibiotic susceptibility analysis of *Enterococcus* spp. isolated from urine. *Przegl Lek.* 2004;61(5):473-6.
12. Kau AL, Martin SM, Lyon W, Hayes E, Caparon MG, Hultgren SJ. Enterococcus faecalis Tropism for the Kidneys in the Urinary Tract of C57BL/6J Mice . *Infection and Immunity*. 2005;73(4):2461-8.
13. Sood S, Malhotra M, Das BK, Kapil A. Enterococcal infections & antimicrobial resistance. *Indian J Med Res* 2008; 128:111-21.

Copyright: © the author(s) and publisher IJM RP. This is an open access article distributed under the terms of the Creative Commons Attribution Non-commercial License, which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite the article: Chaman Lal Girdhar, Naresh Jyoti. Evaluation of Antimicrobial Resistance and Bacterial Spectrum of Infections at a Tertiary Care Teaching Hospital: A Retrospective Analysis. *Int J Med Res Prof.* 2015, 1(3); 172-75.